

Prediction of gametic variance and its use in breeding programs

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Abstract

With genomic selection, gametic variance has been of increasing interest for breeding top animals. Two gametic variance estimates based on SNP effects estimated in 2018, were obtained deterministically or by simulating progeny. They were compared with each other, with the variance of the genomic breeding values of real progeny in fall 2020, and with the variability of the phenotypes of the female offspring. As expected, both predictors of gametic variances were similar, provided that the number of simulated progeny is large enough (500). The correlation between predicted and observed gametic variances was rather high, although limited by the number of offspring available. The correlation between the gametic variance of the sires and the phenotypic variance was weaker, from 0.14 to 0.85 (mean=0.33) according to the heritability of the trait. Gametic variance is useful to select parents of future bulls, but its potential to control phenotypic variability is more limited.

Introduction

Since its implementation in 2009, genomic selection has had a tremendous development, all bulls are genomically evaluated and more and more females are genotyped annually. Young bulls are selected on the basis of their genomic values among a large number of male progeny of bull sires and bull dams. Under the polygenic model, gametic variances of each parent or, equivalently, the Mendelian sampling (MS) variance of the progeny, depend only on inbreeding of the parents. In genomic selection, not only the MS term of the progeny can be predicted but also the gametic variance of each parent, provided that SNP effects and genotype phases of the parents are known. According to the large investments engaged to procreate, evaluate the young bulls (matings, genotyping, embryo production and transfer...) and select the very best ones, it is essential to maximize the probability to generate and find them. This probability depends not only on the pedigree index of the parents but also on their gametic variance. As already shown in the literature, gametic variance is of major importance when breeding top animals (Bijma et al., 2020; Segelke et al., 2014). However, in spite of its increasing interest, little investigation was done on its practical prediction and use (Cole et al., 2011). In this study, we (1) compared two approaches to predict the gametic variance of the genotyped parents; (2) investigated the impact of gametic variance in the selection success of candidates; (3) compared the predicted gametic variances with the variance of genomic breeding values of real genotyped progeny, and (4) with the phenotypic variability of the daughters. In this validation procedure, SNP effects were estimated in Holstein with the information available in June 2018 and genomic breeding values and phenotypes of progeny were those available in fall 2021.

Materials & Methods

Population studied.

We used genotypes and breeding values for 426 widely used bulls in Holstein breeds born between 1991 and 2016. Bulls were selected to have more than 200 daughters with morphology phenotypes, 1000 daughters with lactation phenotypes and 500 genotyped progeny. Genotypes of sires were obtained with different Illumina chip platforms with at least 50K SNPs, and imputed and phased with Fimpute3 software (Sargolzaei et al., 2014).

SNP effects were obtained from the French national genomic evaluation (Boichard et al, 2012) computed in June 2018. These effects were used as input for simulation software and gametic variability prediction. Five production and 10 morphological traits were studied.

Simulation procedure and prediction of gametic variance.

Two different approaches were used to predict the gametic variance of each bull. The first one was based on simulation (Segelke et al, 2014). For each bull mated to one simulated fully homozygous dam, 500 hundred progeny were simulated. For each progeny, segregations were randomly sampled and recombinations were simulated assuming 1cM corresponds to 1 Mbase. Progeny direct genomic value was obtained by combining the SNP effects with its genotypes. The MS variance was estimated by the variance of the DGV of the 500 progeny. Of course, because SNP effects are estimated with an accuracy lower than 1, the MS variance was underestimated.

The second approach is the one described by Santos et al (2019). The gametic variance (V_{theo}) was predicted from the contrasts within loci with the following formula:

$$V_{\text{theo}} = 0.25 [\sum_i \alpha_i^2 + 2 \sum_i \sum_{j>i} \alpha_i \alpha_j (1-2r_{ij})]$$

where : α_i is the contrast between effects of the paternal and maternal alleles for marker i, and r_{ij} is the recombination rate between markers i and j. Recombination rate was assumed equal to 0.5 across chromosomes.

Real data were used to validate gametic variance predictions. For each sire, we computed the variance (V_{obs}) of DGV of progeny genotyped after 2018. The phenotypic variance (V_{yd}) was obtained by computing the variance of the performances adjusted for fixed effects of the evaluation model, *ie* the so-called yield deviation (YD) of the female offspring. For this validation step, both DGV and YD were obtained from the French National evaluation run computed in November 2021. Due to the increase in reference population size and change in French evaluation model (especially change in imputation results due to switch from UMD3 to ARS-UCD1 assembly), markers and haplotypes effects might differ between June 2018 and November 2021, allowing us to test the robustness of our predictors.

Results

Comparison of gametic variance predictions.

The predicted and simulation based gametic variances were very similar, with the same mean, very close standard deviations, and correlations in the [0.82 – 0.94] interval. This correlation lower than one can be attributed to the number of simulated progeny, limited to 500. Therefore in practice, the deterministic approach is preferable because it is more accurate and faster to compute. But simulations can be a more flexible tool especially to study covariations between traits or more complex genetic determinisms with dominance or epistasis.

The same indicators computed with 2021 SNP estimates were consistent with those of 2018, showing a good conservation of SNP effect: the correlations between V_{theo} in 2018 and 2021 ranged from 0.87 to 0.99 according to traits. Correlations are presented on Table 2.

Heterogeneity of gametic variance between bulls.

Gametic variances observed on real progeny DGVs (V_{obs}) varied widely between sires, with a range of 3 to 4 for most traits. This is much more than expected under the polygenic model. Figure 1 shows the distribution of V_{obs} for fat content (left) and stature (right). Differences were particularly large for fat content due to the effect of DGAT1 major gene, and lower but still important for stature with most values ranging from 0.3 to 0.9. This large variability of gametic variance observed across animals highlights its interest when breeding top animals.

Assuming these predictions are accurate and reflect true gametic variance (this depends on the accuracy of SNP effects), very good predictions of extreme deciles can be obtained, as well as the probability to obtain an elite progeny above a given threshold. For instance, for milk yield, the prediction of the 90th percentile was: $P90\% = 0.999 \text{ Genomic Pedigree Index} + 1.29 \sigma_{MS}$ ($R^2 = 0.99$), with σ_{MS} the MS standard deviation for a given couple of parents.

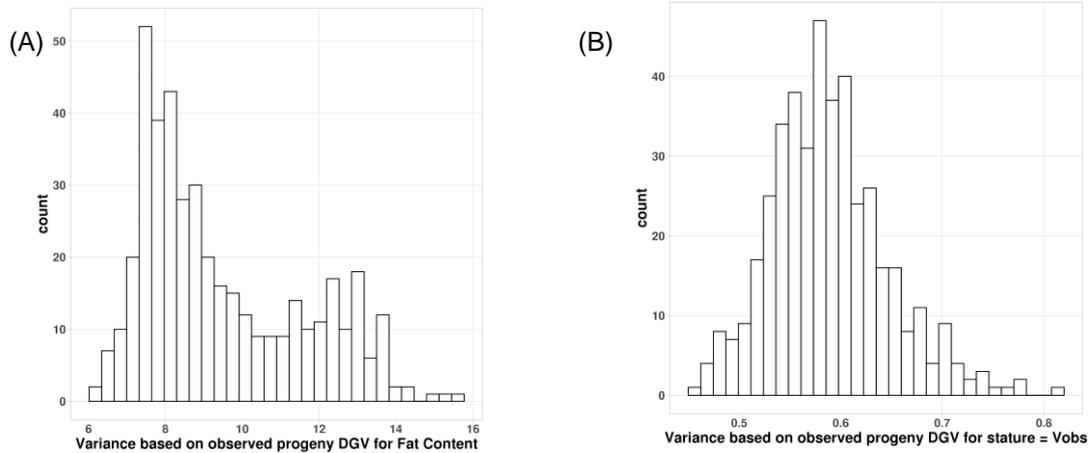


Figure 1. Distribution of progeny DGV variances (V_{obs}) for fat content (A) and stature (B)

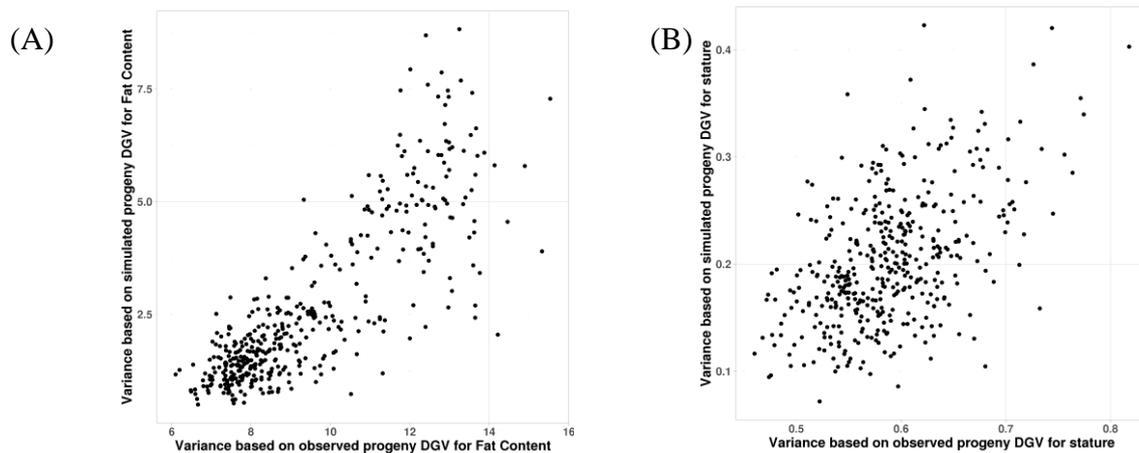


Figure 2. Relationship between within sire variances for simulated and observed progeny, for fat content (A) and stature (B)

Validation of predicted gametic variance

Depending on the traits, the correlations between predicted variance V_{the} in 2018 and observed variance V_{obs} in 2021 ranged from 0.47 to 0.92 and reached 0.62 on average. These moderate values probably reflect both the limited number of progeny and the variability due to dams.

The correlation between V_{theo} and the within bull variance of phenotypes was quite low with an average of 0.25 across traits. On the youngest animal (58 bulls born after 2015), correlations were a bit higher with an average of 0.33. Except for an high correlation for fat content (0.85), correlations were moderate for highly heritable traits (0.44 for protein yield or 0.38 for body depth) and as low as 0.14 for low heritability traits such as rear udder height. All correlations are presented on table 2.

Discussion

This study in Holstein cattle breed investigates the opportunity to predict progeny variability from genotypes and SNP effects.

We observed large variation in progeny variability among bulls supporting the growing interest for an indicator of gametic variability, especially in the choice of bull sires and dams to maximize the probability to conceive top ranking bulls. Validation on real data showed that the proposed theoretical indicator performed quite well to predict the genetic variability of progeny but was not accurate enough to predict the phenotypic performances, highly dependent on environmental factors. Computation of the theoretical predictor is fast, meaning that it can be obtained routinely and used to select animals for the breeding scheme. In this perspective, development of specific selection criteria such as those developed by Bijma et al. (2020) is expected on a routine basis. In spite of its cost of computation, the estimation of gametic variance by simulation of progeny may also be interesting especially in complex situations, mainly in case of non-additive determinism.

Table 2. Correlation between theoretical gametic variance computed in 2018 and others indicators of gametic variability.

Traits	V _{sim_Nov21}	V _{sim_June18}	V _{theo_Nov21}	V _{obs_Nov21}	V _{yd_Nov21}
Protein Yield	0.76	0.91	0.87	0.42	0.44
Fat Yield	0.94	0.94	0.98	0.77	0.16
Protein content	0.89	0.88	0.99	0.84	0.46
Fat Content	0.95	0.95	1.00	0.93	0.85
Milk	0.84	0.90	0.97	0.73	0.15
Udder support	0.77	0.87	0.88	0.59	0.17
Udder Depth	0.78	0.84	0.93	0.37	0.35
Fore Udder Attachment	0.88	0.94	0.95	0.63	0.23
Rear Udder Height	0.88	0.91	0.92	0.55	0.14
Fore Teat Distance	0.89	0.90	0.95	0.64	0.32
Stature	0.83	0.80	0.94	0.64	0.47
Body depth	0.85	0.90	0.94	0.71	0.38
Width at ischium	0.92	0.96	0.96	0.66	0.27
Locomotion	0.83	0.87	0.89	0.51	0.24
Body Condition Score	0.83	0.93	0.94	0.72	0.35
Average	0.86	0.90	0.94	0.65	0.33

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References

- Bijma P., Wientjes Y.C.J., and Calus M.P.L (2020) *Genetics*. 214 (1). 91-107
<https://doi.org/10.1534/genetics.119.302643>
- Sargolzaei M., Chesnais J., and Schenkel F. (2014) *BMC Genomics* 15(1):478.
<http://doi.org/10.1186/1471-2164-15-478>
- Cole J.B and P.M. VanRaden (2011) *J. Anim. Breed. Genet.* 128: 446–455.
<https://doi.org/10.1111/j.1439-0388.2011.00922.x>
- Boichard D., Guillaume F., Baur A., Croiseau P., Rossignol M. N. et al. (2012) *Anim Prod Sci* 52. 115-120. <https://doi.org/10.1071/AN11119>
- Segelke D., Reinhardt F., Liu Z. et al. (2014) *Genet Sel Evol* 46. 42.
<https://doi.org/10.1186/1297-9686-46-42>

Santos D.J.A., Cole J.B., Lawlor T.J.Jr., VanRaden P.M., Tonhati H., et al (2019). J Dairy Sci 102(6) 5279-5294. <https://doi.org/10.3168/jds.2018-15971>